

Table I—Radiation Protection by Thioureas

Compound	Test System	Radiation Dose	Protection	Ref.
Thiourea	Mice	875 rad	Increased lifespan	31
Methylthiourea	Mice	650-700 rad	No effect on lifespan	32
Allylthiourea	Mice	700 rad	No effect on lifespan	8
Phenylthiourea	Mice	650-700 rad	No effect on lifespan	32
α -Naphthylthiourea	Mice	200 rad	No effect on lifespan	33
Ethyl isothiourea	Mice	800 rad	Increased lifespan	34
Sulfanylthiourea	Mice	900 rad	Increased lifespan	35
Guanylthiourea	Mice	900 rad	Increased lifespan	35
Thiourea	Human erythrocytes	84 krad	Protection	12
Thiourea	T ₂ phage	1000 rad/s	Protection	13
Thiourea	Ehrlich ascites cells	4000 rad	Protection	14
Allylthiourea	Human erythrocytes	84 krad	Protection	12

Table II—Ionization Constants ^a

Compound	pK _a
Thiourea	2.54 ^b
<i>N</i> -Methylthiourea	3.06
<i>N,N'</i> -Dimethylthiourea	2.98
<i>N</i> -Methyl- <i>N'</i> -phenethylthiourea	3.12
<i>N</i> -Methyl- <i>N'</i> -phenyl-2-propylthiourea	3.04
2-Imidazolidinethione	3.08
2-Thiobarbituric acid	4.15
Thioacetamide	3.34

^a Determined in 95% ethanol at 25°C. ^b Literature value (H₂O) is 2.03: T. J. Lane, J. A. Ryan, and J. L. Walter, *J. Am. Chem. Soc.*, **78**, 5560 (1956).

presence of 0.0005 mol of divalent metal salt or 0.00033 mol of trivalent metal salt.

Volumes of 50 mL of the same quantities of the metal salts were also titrated with 0.01 M KOH. The pH readings were recorded 2 min after each addition of titrant to allow equilibrium to be reached. Solvent concentration at the end of the titrations (if all 10 additions were made) was ~72% ethanol.

Calculations were performed as previously described (17) with a computer. The log *K* values obtained for the divalent metal complexes are recorded in Table III; values for the trivalent metal complexes are recorded in Table IV. Values for *K*₁, *K*₂, and *K*₃ were obtained from Eqs. 1-3, according to Flood and Loras (22) and Albert (23):

$$K_1 = \frac{\bar{n}}{(1 - \bar{n}) [L^-]} \quad (\text{Eq. 1})$$

$$K_2 = \frac{(\bar{n} - 1)}{(2 - \bar{n}) [L^-]} \quad (\text{Eq. 2})$$

$$K_3 = \frac{(\bar{n} - 2)}{(3 - \bar{n}) [L^-]} \quad (\text{Eq. 3})$$

where \bar{n} is the average number of ligand molecules bound by a metal ion at any stage in complex formation and $[L^-]$ is the concentration of the free chelating species.

Table III—Stability Constants for Cu(II) and Ni(II) Complexes (25°C)

Compound	Cu(II) Complexes ^a			Ni(II) Complexes		
	log <i>K</i> ₁	log <i>K</i> ₂	log β_2	log <i>K</i> ₁	log <i>K</i> ₂	log β_2
Thiourea	1.38 ^b			1.00		
<i>N</i> -Methylthiourea	1.67			1.39		
<i>N,N'</i> -Dimethylthiourea	1.35			1.33		
<i>N</i> -Methyl- <i>N'</i> -phenethylthiourea	1.51			1.47		
<i>N</i> -Methyl- <i>N'</i> -phenyl-2-propylthiourea	1.44			1.45		
2-Imidazolidinethione	1.66			1.40		
2-Thiobarbituric acid	4.35	3.79	8.14	3.76	3.18	6.94
Thioacetamide	2.30			1.69		

^a The product of Cu(II) and thiourea is claimed to be a Cu(I) complex: E. I. Onstott and H. A. Laitinen, *J. Am. Chem. Soc.*, **72**, 4724 (1950). ^b The log β_2 for Cu(II) and thiourea was estimated by Bjerrum to be ~2: L. G. Silén and A. E. Martell, "Stability Constants of Metal-Ion Complexes," The Chemical Society, London, 1964, p. 359.

Formation curves were plotted (\bar{n} versus $-\log L^-$) to show whether stepwise complexation may have taken place. No steps were shown in the plots, probably because of the closeness of the log *K* values.

RESULTS AND DISCUSSION

The *K*₁ values for the Al(III) and Fe(III) complexes were not uncovered, with the exception of the ferric complex of thioacetamide, possibly because of lack of stepwise complexation (no values for \bar{n} below 1 were obtained). No values for the ferric complex of 2-thiobarbituric acid were obtained because of precipitate formation at the beginning of the titration. Values for *K*₂ for the Cu(II) and Ni(II) complexes were not obtained because of the formation of precipitates, with the exception of the complexes of 2-thiobarbituric acid. Although log β values could not be obtained in most cases, for the purposes of comparison and for determining whether thiourea complexes are capable of existence in the presence of cellular complexing agents, the constants found should suffice.

The sequence of stability constants of thiourea, *N*-methylthiourea, and *N,N'*-dimethylthiourea allow some conclusions regarding structure of the complexes. The complexes of *N*-methylthiourea showed decreased stability constants, compared with those of thiourea, for the aluminum and ferric systems, but increased constants for the cupric and nickel systems. For the latter systems, where only *K*₁ values were observed, steric effects of the methyl group would be minimal. For the aluminum and ferric systems, the decrease in stability constants may be attributed to steric effects, since 2:1 and 3:1 complexes are involved. These sequences suggest that both the sulfur and a nitrogen atom are involved in bond formation of the complexes.

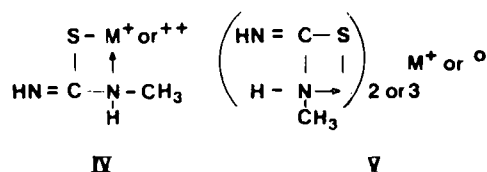
With the complexes of *N,N'*-dimethylthiourea, the additional methyl group caused a decrease in constants for all four metal ions observed. Two methyl groups apparently cause steric hindrance of the 1:1 complexes as well. The longer-chain substituents gave somewhat higher constants than the methyl groups, so the α -methylene groups were less interfering than the methyl groups. For the cyclic thioureas, particularly 2-thiobarbituric acid, where steric hindrance would be less than for the open-chain complexes, the stability constants were the largest of the series. It is also possible that some degree of aromaticity is possible with 2-thiobarbituric acid.

Table IV—Stability Constants for Al(III) and Fe(III) Complexes (25°C)

Compound	Al(III) Complexes		Fe(III) Complexes			
	log K_2	log K_3	log K_1	log K_2	log K_3	log β_3
Thiourea	3.01	1.69		3.55	2.16	
<i>N</i> -Methylthiourea	2.73	1.90		3.00	2.16	
<i>N,N'</i> -Dimethylthiourea	2.61	1.81		2.87	2.28	
<i>N</i> -Methyl- <i>N'</i> -phenethylthiourea	3.79	2.21		3.59	2.41	
<i>N</i> -Methyl- <i>N'</i> -phenyl-2-propylthiourea	3.59	2.21			2.84	
2-Imidazolidinethione	3.61	2.12		3.92	2.40	
2-Thiobarbituric acid	5.15	3.29				
Thioacetamide	3.06	2.23	4.12	3.40	2.25	9.77

It has been shown previously (17) that stability constants for metal complexes of 2-mercaptoimidazoles and mercaptopyrimidines were relatively high, and that both sulfur and nitrogen were involved in bond formation to give four-membered rings. Four-membered metal chelate rings have also been reported for dithiocarbamates and xanthates (24), and for dithiocarboxylates (25). Ruthenium complexes of thioureas have also been postulated to involve the sulfur and nitrogen in four-membered rings (26).

If complexation to sulfur alone were involved, the presence of *N*-methyl substituents would be expected to increase stability constants by electron release. But the probable effect of steric hindrance by the methyl groups, as indicated for the decreased stability of 2:1 and 3:1 Al(III) and Fe(III) complexes, allow the postulation of four-membered chelate rings involving bonding to both sulfur and nitrogen (IV and V).



The magnitude of the metal-binding stability constants observed is somewhat lower than that for simple peptides. Log K_1 values reported for the Cu(II) and Ni(II) complexes of glycyl-DL-alanine, for instance, are 5.92 and 4.08, respectively (27). Although metal complexes of thioureas, with the possible exception of cyclic thioureas, should not be expected to exist in the presence of cellular peptides for any appreciable period, they should be capable of binding to the metal constituents of metalloenzymes.

Wheeler and Ribot (28) observed the protective effect of thiourea and methylthioureas for radiation damage to a synthetic polymer. Presence of the methyl groups distinctly lowered the protective ability of thiourea. It would not appear in this system that metal ions are involved in the radiation damage, which must be due largely to the radicals resulting from radiolysis of water. In this case, the ability of the thioureas to act similarly to the thiol radiation-protectors, in transferring hydrogen atoms to the radiation-produced radicals (29), would appear to offer a more probable mechanism of protection. The ability of thioureas to enter a thione-thiol equilibrium would make hydrogen atom transfer a realistic possibility, and in view of the rather low metal-binding stability constants for thioureas, a more likely mechanism than the ability to bind copper or iron ions, which catalyze cellular oxidations. A previous attempt to relate metal-binding ability to radiation protection of a series of aminoalkyl disulfides and thiosulfates did not give a positive correlation (30).

Involvement of thioureas with metalloenzymes, such as dopamine- β -oxidase, nitrate reductase, or polyphenol oxidase, is still a likely possibility by complexation of the metal constituent, however. A correlation between metal-binding ability and the antimicrobial effects of cyclic thioureas has already been observed (17), possibly through the inhibition of metalloenzyme activity.

REFERENCES

(1) S. Friedman and S. Kaufman, *J. Biol. Chem.*, **240**, 4763 (1965).
 (2) G. A. Johnson, S. J. Boukma, and E. G. Kim, *J. Pharmacol. Exp. Ther.*, **168**, 229 (1969).
 (3) M. J. Seven and L. A. Johnson, "Metal-Binding in Medicine," J. B. Lippincott, Philadelphia, Pa., 1960, p. 321.
 (4) S. Garattini and A. Leonardi, *Giorn. Ital. Chemterap.*, **2**, 18 (1955).

(5) A. K. Sijpesteijn, M. J. Janssen, and G. J. VanderKerk, *Biochim. Biophys. Acta*, **23**, 550 (1957).
 (6) R. H. Garrett and P. Greenbaum, *Biochim. Biophys. Acta*, **302**, 33 (1973).
 (7) H. Less, *Nature (London)*, **158**, 97 (1946).
 (8) P. Alexander, Z. M. Bacq, S. F. Cousens, M. Fox, A. Herve, and J. Lazar, *Radiat. Res.*, **2**, 392 (1955).
 (9) M. M. Jones, *Nature (London)*, **185**, 96 (1960).
 (10) W. O. Foye and J. Mickles, "Progress in Biochemical Pharmacology," vol. 1, Butterworths, Washington, D.C., 1965, pp. 152-160.
 (11) H. Langendorff, R. Koch, and U. Hagen, *Arch. Intern. Pharmacodyn.*, **100**, 1 (1954).
 (12) K. Flemming, *Folia Haemat. (Leipzig)*, **77**, 147 (1960); through *Chem. Abstr.*, **55**, 3687 (1961).
 (13) P. Howard-Flanders and P. Jockey, *Int. J. Radiat. Biol.*, **2**, 361 (1960).
 (14) V. Drasil, *Biol. Eff. Ioniz. Radiat. Mol. Level, Proc. Symp.*, 1962, **91-7**; through *Chem. Abstr.*, **58**, 14413, (1963).
 (15) A. Charlesby and P. M. Kopp, *Proc. Roy. Soc., Ser. A*, **291**, (1424), 113 (1966).
 (16) L. G. Sillén and A. E. Martell, "Stability Constants of Metal-Ion Complexes," Suppl. 1, The Chemical Society, London, 1971, pp. 238-240.
 (17) W. O. Foye and J.-R. Lo, *J. Pharm. Sci.*, **61**, 1209 (1972).
 (18) D. M. G. Armstrong, *Chem. Ind. (London)*, **1955**, 1405.
 (19) W. O. Foye and J. C. Anderson, *J. Pharm. Sci.*, **58**, 1558 (1969).
 (20) W. O. Foye and J. N. Sane, *J. Pharm. Sci.*, **66**, 923 (1977).
 (21) A. Albert and E. P. Serjeant, "The Determination of Ionization Constants," Chapman and Hall, London, 1971.
 (22) H. Flood and V. Loras, *Tidsskr. Kjem. Bergv. Met.*, **4**, 35 (1944); through *Chem. Abstr.*, **41**, 6488 (1947).
 (23) A. Albert, *Biochem. J.*, **47**, 531 (1950).
 (24) J. Chatt, L. A. Duncanson, and L. M. Venanzi, *Nature (London)*, **177**, 1042 (1956).
 (25) B. G. Werden, E. Billig, and H. B. Gray, *Inorg. Chem.*, **5**, 78 (1966).
 (26) R. P. Yaffe and A. F. Voigt, *J. Am. Chem. Soc.*, **74**, 2503 (1952).
 (27) L. G. Sillén and A. E. Martell, "Stability Constants of Metal-Ion Complexes," Suppl. 1, The Chemical Society, London, 1971, p. 386.
 (28) O. H. Wheeler and R. A. Ribot, *Chem. Ind. (London)*, **30**, 1017 (1969).
 (29) W. O. Foye, *Int. J. Sulfur Chem.*, **8**, 161 (1973).
 (30) W. O. Foye and J.-M. Hu, *J. Pharm. Sci.*, **68**, 202 (1979).
 (31) R. H. Mole, J. St. L. Philpot, and J. R. V. Hodges, *Nature (London)*, **166**, 515 (1950).
 (32) L. F. Semenov and E. A. Prokudina, *Med. Radiol. (Moscow)*, **1/4**, 70 (1956); through *Chem. Abstr.*, **51**, 13212 (1957).
 (33) H. Langendorff, R. Koch, and U. Hagen, *Arch. Intern. Pharmacodyn.*, **104**, 57 (1955).
 (34) J. F. Noble, V. J. Plzak, R. M. Dowben, and J. Doull, *Fed. Proc. Fed. Am. Soc. Exp. Biol.*, **16**, 325 (1957).
 (35) K. Stratton and E. M. Davis, *Int. J. Radiat. Biol.*, **5**, 105 (1962).

ACKNOWLEDGMENTS

Abstracted in part from a thesis submitted by C.-C. Chao to the Massachusetts College of Pharmacy and Allied Health Sciences in partial fulfillment of the Master of Science degree requirements. Part XVIII of this series is W. O. Foye and J.-M. Hu, *J. Pharm. Sci.*, **68**, 202 (1979).